E. coli growth media

LB:
For 1 L
• 10 g tryptone
• 5 g yeast extract
• 10 g NaCl
• Optional: Bring up the volume to around 900 mL with ddH2O, then adjust the pH to 7.4
• Bring up the volume to 1 L with ddH2O
• Sterilize by autoclaving for 30 min
To make LB-agar, add 15 g of agar prior to autoclaving

Low-salt LB:
For 1 L
• 10 g tryptone
• 5 g yeast extract
• 5 g NaCl
• Optional: Bring up the volume to around 900 mL with ddH2O, then adjust the pH to 7.4
• Bring up the volume to 1 L with ddH2O
• Sterilize by autoclaving for 30 min
To make low-salt LB-agar, add 15 g of agar prior to autoclaving

SOC:
For 1 L
• 20 g tryptone
• 5 g yeast extract
• 0.5 g NaCl
• 0.186 g KCl
• 0.952 g MgCl₂
• Bring up the volume to around 900 mL with ddH2O, then adjust the pH to 7.4 with 10 N NaOH
• Bring up the volume to 1 L with ddH2O
• Sterilize by autoclaving for 30 min
• Add 20 mL of sterile 1 M glucose immediately before use

1 M glucose:
For 100 mL
• Dissolve 18 g glucose in 100 mL ddH₂O
• Sterilize by filtering through 0.2 µm or by autoclaving for 15 min

Antibiotic stocks

Ampicillin (1000x):
• Dissolve 5 g ampicillin in 25 mL ddH₂O
• Add 25 mL absolute ethanol
• Store at -20 °C
Carbenicillin (1000x for liquid media, 2000x for plates):
• Dissolve 2.5 g carbenicillin in 25 mL ddH₂O
• Add 25 mL absolute ethanol
• Store at -20 °C

Chloramphenicol (1000x):
• Dissolve 1.7 g chloramphenicol in 50 mL absolute ethanol
• Store at -20 °C

Kanamycin (1000x, but 250x for autoinduction media):
• Dissolve 1.25 g ampicillin in 50 mL ddH₂O
• Sterilize by filtering through 0.2 µ
• Store at -20 °C in 1.5-mL aliquots

Tetracycline (1000x):
• Dissolve 0.75 g tetracycline in 50 mL absolute ethanol or isopropanol
• Store at -20 °C in 1.5-mL aliquots